



Understanding the Biochemical Basis of Resistance in Rice to Leaf folder (*Cnaphalocrocis medinalis*)

Anil Varma Nalla ^{a*}, S. Nadaradjan ^a, D. Adiroubane ^a
and K. Kumar ^a

^a Pandit Jawaharlal Nehru College of Agriculture and Research Institute, Karaikal-609603, U.T. of Puducherry, India.

Authors' contributions

This work was carried out in collaboration among all authors. All authors read and approved the final manuscript.

Article Information

DOI: 10.9734/IJPSS/2023/v35i153088

Open Peer Review History:

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://www.sdiarticle5.com/review-history/99796>

Original Research Article

Received: 12/03/2023
Accepted: 17/05/2023
Published: 10/06/2023

ABSTRACT

Rice leaf folder has been recorded to cause major loss in rice yield. To know the relevance of biochemical factors in conferring tolerance to rice leaf folder *Cnaphalocrocis medinalis*, a study was conducted utilizing 196 rice accessions at Pandit Jawaharlal Nehru College of Agriculture and Research Institute, Karaikal. Standard evaluation system developed by IRRI for leaf folder complex was followed for screening the rice accessions and entries were ranked accordingly based on the leaf damage. Among these entries, top five entries along with the susceptible check (TN1) were selected for analysis of biochemical factors such as total chlorophyll, total sugars, reducing sugars, total phenols, total soluble protein and proline. Results revealed that high level of total phenols, moderate chlorophyll content, and low sugar content in leaves conferred resistance to this pest in rice.

*Corresponding author: E-mail: anilvarmanalla@gmail.com;

Keywords: Total sugars; reducing sugars; phenol content; total soluble protein; proline.

1. INTRODUCTION

The rice leaffolder *Cnaphalocrocis medinalis* (Guenee) is one of the destructive insect pests of rice. It was considered as minor or sporadic pest in the past in many Asian countries. Due to this pest, loss in seed yield have been reported from 20 per cent at Tillering stage to as high as 50 per cent at Flowering stage [1,2]. Extensive feeding results in reduction in photosynthetic ability, vigour and predisposes the leaves to bacterial and fungal infection [3]. In certain cases it has been recorded to cause 63 to 80 per cent loss in rice yield [4]. Intensive and extensive cultivation of rice for maximization of yield and use of new strategies in agriculture, have led to complete changeover in the ecology of rice field and because of the changed agro ecosystem, some minor pests have attained major status.

The mechanism of resistance may be due to physical, biochemical or both combined factors. Accumulation of defense enzymes, chemicals and resistant proteins by insect feeding has been reported in many insect-plant interactions [5]. The plant strategy to deter feeding herbivore has become an important aspect of insect-plant interaction studies and is gaining tremendous importance. Among the plant chemicals, presence of increased or decreased amount of both the nutritional compounds and non nutritional secondary substances influences the resistance or susceptibility of plants to insects [6]. According to Kogan and Paxton [7], many changes that occur following herbivory result in accumulation of phenolic compounds. Also, the direct defense is known to reduce insect growth rates by interfering with the digestibility and nutritional quality of plant tissues [8]. To identify the resistance genotypes in rice to leaf folder complex, 196 entries were taken for conducting the field trial at Karaikal. Resistant entries were selected based on the screening procedure (IRRI standard evaluation procedure). To know the relevance of biochemical factors responsible for the resistance mechanism in these genotypes, biochemical analysis such as total chlorophyll, total sugars, reducing sugars, total phenols, total soluble protein and proline were carried out. Biochemical studies of rice varieties will be helpful in confirming the physiological antibiosis of the new rice germplasm. Feeding activities of herbivorous insects often result in physiological, morphological and chemical changes in the form of accumulation of the compounds having

defensive properties. The biochemical factors are chemicals that affect insect behaviour, physiology and growth. Some biochemical factors are associated with repellence, feeding deterrence toxicity or adverse effects on insects [9].

2. MATERIALS AND METHODS

Rice accessions have been received from the Indian Institute of Rice research (IIRR) Hyderabad with two susceptible check (TN 1). Standard evaluation system was followed for screening the rice accessions developed by IRRI for leaf folder complex such as *Cnaphalocrocis medinalis*, *Marasmia patnalis*, *M. ruralis* and *M. exigua*. Standing water was maintained continuous to a height of 2 to 5 cm throughout the crop season by irrigating the field on need basis. Fertilizers viz., N: P₂O₅:K₂O were applied @ 120:60:60 kg/ha in the form of Urea, Diammonium phosphate (DAP) and Muriate of potash (MOP) respectively. Full doses of P₂O₅ and K₂O along with half dose of N were applied before transplanting as a basal application, while remaining half dose of N was applied in two equal splits at tillering and panicle initiation stages of the crop. Weeds were manually removed from experimental field to avoid crop-weed competition during crop period.

After screening the accessions, the resistant entries with least mean leaf folder damage were identified. Top five entries based on the ranking and a susceptible entry TN 1 was taken for biochemical analysis. To determine the biochemical factors responsible for imparting resistance in the promising genotypes, estimation of total chlorophyll, total sugars, reducing sugars, total phenols, protein and proline were carried out.

2.1 Estimation of Biochemical Factors

The biochemical factors were estimated from the leaf samples. Total chlorophyll was estimated following Hiscox and Israelstam. For total and reducing sugars Nelson-Somogyi method was followed (Eric Fournier, 2001), while total phenol was estimated following Sadasivam and Manikkam (1996). For estimation of protein, Lowry's method was followed [10] and proline was estimated employing Bates et al. [11].

2.2 Statistical Analysis

Data on biochemical parameters analyzed using AGRES software for its significance.

3. RESULTS AND DISCUSSION

Since leaf folder complex feeds on the leaves, to find out whether the intensity of greenness has a role on attracting the pest, total chlorophyll content was analyzed in the selected resistant entries along with a susceptible check. Results implied that susceptible entry TN 1 has 4.83 mg/g while the resistant entries having comparatively less amounts of chlorophyll (Table 1). Similar type of result was obtained by Xu et al. against the leaf folder incidence. This result suggests that greenness has an influence in attracting leaf folder pest.

As leaf folder complex is a chewing pest, to know the role of sugars on the palatability of leaves, total reducing sugars was estimated from the leaves of resistant and susceptible entries.

Significant variation was observed among the tested entries for total sugars. Total sugars were higher in the susceptible entry TN 1 (129.86 mg/g) than resistant ones (Table 2). These findings were in accordance with the Nanda et al., [12]; Padhi, [13]; Chandramani et al., [14]. Higher amounts of total sugars were reported in brown planthopper susceptible entries, Tellahamsa and Jaya [15]. *Sogatella furcifera* populations were positively correlated with the total sugars and amino acids [16].

Amount of reducing sugars also influenced the incidence of pest. Amount of reducing sugars in resistant entries ranged from 27.20 to 56.02 mg/g and the susceptible check TN 1 has 53.57 mg/g. Among the resistant entries, UPR 3506-7-1-1 showed 56.02 mg/g which was higher than the susceptible check. The cause of resistance may be due to high phenolic content (15.66 mg/g) (Table 3). Similar results were reported by Nanda et al., [12]; Padhi [13], Chandramani et al., [14] and Ashrith et al., [17].

Table 1. Total chlorophyll of selected rice genotypes showing differential reaction to rice leaf folder

S. No.	Accession	(%) Leaf damage	Total chlorophyll (mg/g)
1.	ARRH-3626	6.82	2.97
2.	OR 2324-8	6.94	2.21
3.	CR 2698	7.72	2.72
4.	UPR 3506-7-1-1	7.81	3.28
5.	HUBR 10-9	7.81	2.29
6.	R 1528-1058-1-110-1	7.93	3.09
7.	CN 1561-70-19-35-9-MLD 1	7.93	3.37
8.	TN-1	45.82	4.83
Mean	--	--	3.09
C.D (P=0.05)	--	--	0.49
C.V%	--	--	9.08

Table 2. Total sugars of selected rice genotypes showing differential reaction to rice leaf folder

S. No.	Accession	(%) Leaf damage	Total sugars (mg/g)
1.	ARRH-3626	6.82	23.18
2.	OR 2324-8	6.94	51.90
3.	CR 2698	7.72	53.02
4.	UPR 3506-7-1-1	7.81	25.49
5.	HUBR 10-9	7.81	17.96
6.	R 1528-1058-1-110-1	7.93	38.62
7.	CN 1561-70-19-35-9-MLD 1	7.93	26.82
8.	TN-1	45.82	129.86
Mean	--	--	45.86
C.D (P=0.05)	--	--	1.69
C.V%	--	--	2.10

Table 3. Reducing sugars of selected rice genotypes showing differential reaction to rice leaf folder

S. No.	Accession	(%) Leaf damage	Reducing sugars (mg/g)
1.	ARRH-3626	6.82	27.81
2.	OR 2324-8	6.94	47.31
3.	CR 2698	7.72	34.93
4.	UPR 3506-7-1-1	7.81	56.02
5.	HUBR 10-9	7.81	25.46
6.	R 1528-1058-1-110-1	7.93	27.20
7.	CN 1561-70-19-35-9-MLD 1	7.93	34.21
8.	TN-1	45.82	53.57
Mean	--	--	38.31
C.D (P=0.05)	--	--	1.82
C.V%	--	--	2.72

Entries with low amount of phenols were prone to attack by leaf folder larva. Amount of total phenols ranged from 5.67 to 15.66 mg/100g. Susceptible entry TN 1 (5.67 mg/100g) has lesser amount of phenolic compounds (Table 4). These results were in concurrence with the findings of Loka Reddy et al., [18] and Chandramani et al. [14] in brown plant hopper affected leaves. Similar results were reported by Rathika [19] and Ashrith et al. [17] in rice for leaf folder indicating the presence of higher phenols in resistant entries.

Amount of total protein was higher in the entries which were resistant against the leaf folder while the susceptible entry had comparatively lesser amounts. Amount of total protein ranged from 5.80 mg/g to 23.08 mg/g (Table 5). Amount of protein in the susceptible check TN1 was 11.84 mg/g implying that protein content did not influenced the resistance against leaf folder. These results were in contrast to the findings of

Suchita et al. [20] in cotton against Mealybugs that protein content is higher in susceptible entries.

The data from previous studies suggested that proline has regulatory function, controls plant development and act as signal molecules (Laszlo Szabados and Arnould Savoure, 2004). Proline metabolism can also influence programmed cell death in plants. In *Arabidopsis*, incompatible plant-pathogen interactions trigger a hypersensitive response (HR) via reactive oxygen species (ROS) signals, which is accompanied by local activation of *P5CS2* and proline accumulation [21]. Proline was recently proposed to modulate the plant defence response to *Agrobacterium tumefaciens*. Proline accumulates in plant tumours, and functions as a competitive antagonist of gamma-aminobutyric (GABA)-dependent plant defence, interfering with the GABA-induced degradation of quorum-sensing signal [22-28].

Table 4. Total phenols of selected rice genotypes showing differential reaction to rice leaf folder

S. No.	Accession	(%) Leaf damage	Phenols (mg/100 g)
1.	ARRH-3626	6.82	6.89
2.	OR 2324-8	6.94	12.76
3.	CR 2698	7.72	12.49
4.	UPR 3506-7-1-1	7.81	15.66
5.	HUBR 10-9	7.81	9.57
6.	R 1528-1058-1-110-1	7.93	10.49
7.	CN 1561-70-19-35-9-MLD 1	7.93	14.82
8.	TN-1	45.82	5.67
Mean	--	--	10.77
C.D (P=0.05)	--	--	3.74
C.V%	--	--	19.83

Table 5. Total soluble protein of selected rice genotypes showing differential reaction to rice leaf folder

S. No.	Accession	(%) Leaf damage	Protein (mg/g)
1.	ARRH-3626	6.82	5.80
2.	OR 2324-8	6.94	12.85
3.	CR 2698	7.72	20.21
4.	UPR 3506-7-1-1	7.81	23.08
5.	HUBR 10-9	7.81	11.21
6.	R 1528-1058-1-110-1	7.93	11.62
7.	CN 1561-70-19-35-9-MLD 1	7.93	18.44
8.	TN-1	45.82	11.84
Mean	--	--	14.38
C.D (P=0.05)	--	--	0.92
C.V%	--	--	3.6

Table 6. Proline of selected rice genotypes showing differential reaction to rice leaf folder

S. No.	Accession	(%) Leaf damage	Proline (ppm)
1.	ARRH-3626	6.82	33.90
2.	OR 2324-8	6.94	24.95
3.	CR 2698	7.72	52.72
4.	UPR 3506-7-1-1	7.81	36.75
5.	HUBR 10-9	7.81	36.07
6.	R 1528-1058-1-110-1	7.93	47.72
7.	CN 1561-70-19-35-9-MLD 1	7.93	51.65
8.	TN-1	45.82	113.03
Mean	--	--	49.59
C.D (P=0.05)	--	--	9.38
C.V%	--	--	10.80

Very few or nil reports are found for the role of proline against pathogen or pest incidence. In order to investigate the role of proline against pest damage, the proline content was analysed in the rice entries (Table 6). Interestingly the susceptible check TN 1 found to have significantly higher level of proline when compared to resistant entries implying the fact that more damage induce the synthesis of proline which may act as a signal molecule for plant defence mechanism. By further studies this may be proved [29-34].

4. CONCLUSION

By the present study, it is suggested that rice genotypes having high phenolic compounds, moderate chlorophyll content and lower sugar content could be utilized in the breeding programme for developing resistant varieties for leaf folder. Varieties with high phenolic compounds, moderate chlorophyll content and lower sugar content may be recommended for the areas with high leaf folder infestation in rice.

CONFERENCE DISCLAIMER

Some part of this manuscript was previously presented in the conference: 3rd International Conference IAAHAS-2023 "Innovative Approaches in Agriculture, Horticulture & Allied Sciences" on March 29-31, 2023 in SGT University, Gurugram, India. Web Link of the proceeding: <https://wikifarmer.com/event/iaahas-2023-innovative-approaches-in-agriculture-horticulture-allied-sciences/>.

COMPETING INTERESTS

Authors have declared that no competing interests exist.

REFERENCES

1. Manjunatha M, Surivanna BK, Naik M. Field evaluation of Ezeetab (deltamethrin 25% tablets) against rice leaf folder. *Insect Environ.* 2002;8:42.
2. Padmavathi CH, Katti G, Padmakumari AP, Voleti SR, Subba Rao LV. The effect

- of leaffolder *Cnaphalocrocis medinalis* (Guenee) (Lepidoptera: Pyralidae) injury on the plant physiology and yield loss in rice. J Appl Entomol. 2013;137(4):249-56.
3. Sabir AM, Ahmed S, Qadir A. Role of weather on major pest insects of rice crop and their control. Germany: LAP LAMBERT Academic Publishing; 2012.
 4. Rajendran R, Rajendran B, Sundrababu PC. Varietal resistance of rice to leaffolder. Int Rice Res Newsl. 1986;11(4):17-8.
 5. Radja Commare R, Nandakumar R, Kandan A, Suresh S, Bharathi M, Raguchander T et al. Pseudomonas fluorescences based bio-formulation for the management of sheath blight and leaffolder insect in rice. Crop Prot. 2002;21(8):671-7.
 6. Bharathi M. Biochemical basis of resistance in rice to insect pests. In: Ananthkrishnan TN. National Symposium on Biochemical Bases of Host Plant Resistance of Insects. New Delhi; 1996. Indian Agricultural Research Institute. National Academy of agricultural sciences. p. 26-34.
 7. Kogan M, Paxton J. Natural inducers of plant resistance to insects Hedin P A. Plant Resistance to Insects. Am Chem Soc Symp S. 1983;208:153-71.
 8. Johnson R, Narvaez J, An G, Ryan C. Expression of proteinase inhibitors I and II in transgenic tobacco plants: effects on natural defense against *Manduca sexta* larvae. Proc Natl Acad Sci U S A. 1989;86(24):9871-5.
 9. Saxena RC. Biochemical bases of resistance in rice varieties. In: Green MB, Hedin PA, editors. Natural resistance of plants to pests: role of allelochemicals. Washington: American Chemical Society. 1986;142-59.
 10. Lowry OH, Rosebrough NJ, Farr AL, Randall RJ. N. lugens. Indian J Entomol. J. Biol. Chem. 62. 1951;193:265 (The original method):239-41.
 11. Bates LS, Waldren RP, Teare ID. Rapid determination of free proline for water stress studies. Plant Soil. 1973;39(1):205-7.
 12. Nanda UK, Dash D, Rath LK. Biochemical basis of resistance in rice to BPH; 2000.
 13. Padhi G. Biochemical basis of resistance in rice to yellow stem borer, *Scirpophaga incertulas* Wlk. Madras Agricultural Journal. 2004;91(4-6):253-6.
 14. Chandramani P, Rajendran R, Sivasubramanian P, Muthiah C. Management of hoppers in rice through host nutrition- A novel approach. J Biopesticides. 2009;2(1):99-106.
 15. Sujatha G, Reddy GPV, Murthy MMK. Effects of certain biochemical factors on the expression of resistance of rice varieties to brown planthopper (*Nilaparvata lugens* Stal.). J Res APAU. 1987;15:124-8.
 16. Rath LK, Mishra DS, Panda SK. Effect of certain micronutrient fertilizers on the biochemical changes in rice and their correlation with *Sogatella furcifera* Howrath. Ann Plant Prot Sci. 1999;7:8-12.
 17. Ashrith KN, Kalleshwaraswamy MC, kumar D, Dhananjaya CB. Morphological and biochemical basis of resistance in rice genotypes against two major lepidopteran pests. Indian J Entomol. 2020;82(4). DOI: 10.5958/0974 8172.2020.00158.3: 739-44
 18. Loka Reddy K, Pasalu IC, Sreenivasa Raju A, Reddy DDR. Biochemical basis of resistance in rice cultivars to brown plant hopper *Nilaparvata lugens* (Stal.). J Entomol Res. 2004;28(1):79-85.
 19. Rathika M. Studies on the tritrophic interaction of rice leaf folder [M. Sc. Thesis]. Coimbatore: Tamil Nadu Agricultural University; 2008.
 20. Suchita RG, Barkhade UP, Moharil MP, Ugale TB. Biochemical basis of resistance to cotton cultivars against mealy bugs (*Phenacoccus solenopsis* Tinsley). Crop Res. 2011;42:(1, 2 & 3):320-323.
 21. Fabro G, Kovács I, Pavet V, Szabados L, Alvarez ME. Proline accumulation and AtP5CS2 gene activation are induced by plant-pathogen incompatible interactions in *Arabidopsis*. Mol Plant Microbe Interact. 2004;17(4):343-50.
 22. Haudecoeur E, Planamente S, Cirou A, Tannières M, Shelp BJ, Moréra S et al. Proline antagonizes GABA-induced quenching of quorum sensing in *Agrobacterium tumefaciens*. Proc Natl Acad Sci U S A. 2009;106(34):14587-92.
 23. Dhaliwal GS, Shaki HN, Gill PS, Maskina MS. Field reaction of rice varieties to leaf folder at various nitrogen levels. Int Rice Res Newsl. 1979;4(3):7.
 24. Fournier E. Current protocols in food. Anal Chem E1.1.1-E1.1.8. 2001.
 25. Garcia-Olmedo F, Salcedo G, Sanchez-Monge R, Gomez L, Royo J, Carbonero P.

- Oxf Surv Plant Mol Cell Biol. 1987;4:275-334.
26. Hiscox JD, Israelstam GF. A method for the extraction of chlorophyll from leaf tissues without maceration. Can J Bot. 1979;57(12):1332-4.
 27. Khan ZH, Gupta SL, Ramamurthy VV, Dey D. Biodiversity inventory of lepidopterous insects associated with rice agroecosystem. *Shashpa*. 1999;2(107).
 28. Szabados L, Savouré A. Proline: a multifunctional amino acid. Trends Plant Sci. 2010;15(2):89-97.
 29. Padmavathi CH, Katti G, Padmakumari AP, Voleti SR, Subba Rao LV. The effect of leaf folder *Cnaphalocrocis medinalis* (Guenee) (Lepidoptera: Pyralidae) injury on the plant physiology and yield loss in rice. J Appl Entomol. 2013;137(4):249-56.
 30. Sadasivam S, Manikkam A. Biochemical methods. 2nd Ed. New Age International (P) Limited Publishers. 1996;20-21:193-194.
 31. Singh HM, Srivastava RK, Rizvi SMA, Elazegui FA, Castilla NP, Savary S. Yield reduction due to brown spot and leaf folder injuries at varying levels of fertilizers and water supply to rice crop. Ann Plant Prot Sci. 2003;11:16-9.
 32. Xu J, Wang QX, Wu JC. Resistance of cultivated rice varieties to *Cnaphalocrocis medinalis* (Lepidoptera: Pyralidae). J Econ Entomol. 2010;103(4): 1166-71.
 33. Manjunatha M, Surivanna BK, Naik M. Field evaluation of Ezeetab (deltamethrin 25% tablets) against rice leaf folder. Insect Environ. 2002;8:42.
 34. Sabir AM, Ahmed S, Qadir A. Role of weather on major pest insects of rice crop and their control. Germany: LAP Lambert Academic Publishing; 2012.

© 2023 Nalla et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:
<https://www.sdiarticle5.com/review-history/99796>