



# Osmopriming with Polyethylene Glycol (PEG-6000) Improves the Action of Seed Germination, Growth, and Physiology in Carrot

Eshita Kundu <sup>a</sup> and Sanjoy Kumar Bordolui <sup>a\*</sup>

<sup>a</sup> Department of Seed Science and Technology, Bidhan Chandra Krishi Viswavidyalaya, Mohanpur, Nadia, West Bengal, India.

## **Authors' contributions**

This work was carried out in collaboration between both authors. Both authors read and approved the final manuscript.

## **Article Information**

DOI: <https://doi.org/10.9734/ijpss/2025/v37i15293>

## **Open Peer Review History:**

This journal follows the Advanced Open Peer Review policy. Identity of the Reviewers, Editor(s) and additional Reviewers, peer review comments, different versions of the manuscript, comments of the editors, etc are available here: <https://pr.sdiarticle5.com/review-history/130700>

**Original Research Article**

**Received: 24/11/2024**

**Accepted: 29/01/2025**

**Published: 03/02/2025**

## **ABSTRACT**

In order to improve germination and vigour, the current study was conducted using three carrot varieties viz. Carrot Florence (G<sub>1</sub>), Deb Kuroda-1 (G<sub>2</sub>), and Deb Kuroda-3 (G<sub>3</sub>), and various concentrations and durations of PEG-6000, including 0.1 MPa for 24 hours (T<sub>2</sub>), 0.1 MPa for 48 hours (T<sub>3</sub>), 0.25 MPa for 24 hours (T<sub>4</sub>), 0.25 MPa for 48 hours (T<sub>5</sub>), 0.40 MPa for 24 hours (T<sub>6</sub>), and 0.40 MPa for 48 hours (T<sub>7</sub>), non-primed seeds (T<sub>1</sub>). A pre-sowing technique called seed priming produces a physiological environment that promotes more efficient seed germination. The experiment was carried out in the Department of Seed Science and Technology's seed testing lab at the BCKV, Mohanpur, Nadia, West Bengal, India. According to the experiment's results, seeds

\*Corresponding author: E-mail: [sanjoy\\_bordolui@rediffmail.com](mailto:sanjoy_bordolui@rediffmail.com);

treated with 0.25 MPa PEG-6000 soaking for 48 hours produced the best results among treatments over genotype; these seeds showed notably greater potential than seeds treated with other priming concentrations and durations. Deb Kuroda-3 is the best from a germination perspective, and Deb Kuroda-1 is the best from a vigour perspective. The best results were clearly obtained with a 0.25 MPa PEG-6000 soaking duration of 48 hours for seed quality parameters like germination energy (47.273), seedling Vigour Index-I (639.032), and germination index (5.503). Therefore, to improve seedling establishment, PEG-6000 0.25 Mpa pre-sowing treatment for 48 hours is recommended for carrot.

**Keywords:** Germination; PEG-6000; priming; vigour.

## 1. INTRODUCTION

One of the most important vegetable crop in India is carrot (*Daucus carota* L.) (2n=18). This is a biennial plant in the Apiaceae family. Since seed is a key component of crop production, optimal seed germination is a prerequisite for a successful stand establishment. These days, the proportion of seed germination, emergence, and vigour of seedlings has been negatively impacted by many environmental and abiotic stressors, which eventually leads to low crop output. Numerous physiological and non-physiological methods are available to improve seed performance and overcome environmental limitations in order to speed up the germination process. Seed priming is a low-cost effective hydration technique to stimulate seed germination. During priming, seeds go through a physiological process, i.e. controlled hydration and drying which results in enhanced and improved pre-germinative metabolic process for rapid germination. Seed priming can synchronize seed germination, and increase emergence (Heydecker, 1973).

Instead of using pure water, osmopriming entails soaking of seeds in an osmotic solution with a low water potential. The low water potential of osmotic solutions causes water to enter seeds slowly, allowing for progressive imbibition and the activation of early germination phases while preventing radicle protrusion. Various osmotic solutions, including sugar, polyethylene glycol (PEG), glycerol, sorbitol, and mannitol, are used depending on the type of plant, and they are then allowed to air dry before being sown (Slama et al., 2007). Seed priming can improve crop performance under stress conditions, speed up germination, and reduce germination time (Basu, 1976; Chakraborty and Bordolui, 2021).

Benefits of seed priming include improved crop production, maturity, photo and thermo-

dormancy release, nutrient uptake, and water use efficiency (Slama et al., 2007). Therefore, our goal was to ascertain the proper PEG 6000 concentration and duration, which are crucial for carrot seed priming. Given the aforementioned factors, the current study investigated the effects of PEG-6000 seed priming at different doses and periods, along with dry seeds as a control, on vigour status, seedling growth, and germination in a laboratory setting.

## 2. MATERIALS AND METHODS

In the current study, three carrot genotypes and different osmo priming concentrations and durations were used which was carried out during 2022 at the Seed Testing Laboratory, Department of Seed Science and Technology, BCKV, Mohanpur, Nadia, West Bengal, using a completely randomised design with three replications. Three carrot genotypes were Carrot Florence (G<sub>1</sub>), Deb Kuroda-1 (G<sub>2</sub>), Deb Kuroda-3 (G<sub>3</sub>). PEG-6000 was applied at 0.1 MPa for 24 hrs (T<sub>2</sub>), 0.1 MPa for 48 hrs (T<sub>3</sub>), 0.25 MPa for 24 hrs (T<sub>4</sub>), 0.25 MPa for 48 hrs (T<sub>5</sub>), 0.40 MPa for 24 hrs (T<sub>6</sub>), 0.40 MPa for 48 hrs (T<sub>7</sub>). The control (T<sub>1</sub>) was non-primed seeds. AICRP Vegetable provided the seeds, which were analyzed in the Seed Testing Laboratory.

### 2.1 Time to 50% Germination

The number of seeds that germinated each day was noted using the AOSA method. The following formulas from Coolbear et al. (1984), as modified by Farooq et al. (2005), were used to calculate the time of 50% germination (T<sub>50</sub>):

$$T_{50} = t_i + \frac{\left(\frac{N}{2} - n_i\right)(t_j - t_i)}{(n_j - n_i)}$$

Where,

N stands for final number of germination and  $n_i$ ,  $n_j$  are cumulative number of seeds germinated by adjacent counts at times  $t_i$  and  $t_j$  when  $n_i < N/2 < n_j$ .

## 2.2 Mean Germination Time (MGT)

The Ellis and Roberts (1981) equation was used to calculate the mean germination time (MGT):

$$MGT = \frac{\sum Dn}{\sum n}$$

Where D is the number of days measured from the start of germination and n is the number of seeds that germinated on day D.

## 2.3 Germination Percentage

Germination percentage (G) is computed as:

$$G = \frac{X}{Y} \times 100$$

Where X is the number of normal seedlings produced and Y is the total number of seeds taken for germination (ISTA, 1996). Percentage is used to illustrate it.

## 2.4 Germination Index (GI)

According to Ruan et al., (2002), the germination index (GI) was calculated using this formula:

$$GI = \frac{\text{Number of germinated seeds}}{\text{Day of first count}} + \dots + \frac{\text{Number of germinated seeds}}{\text{Day of last count}}$$

## 2.5 Germination Energy

On the fourth day after planting, the germination energy (GE) was noted. In relation to the total number of seeds tested, it is the percentage of seeds that germinated 4 days after planting (Ruan et al., 2002).

## 2.6 Germination Percentage

Cotton was placed in the petridish, and after that blotting paper was placed on it. Then it was wetted by distilled water. After the seeds were prepared, they were put on the blotting paper and covered with a lid. Such eight pairs of

petridish as were kept in the germinator for each genotype and lot. The petridishes were removed from the seed germinator after fourteen days, and the numbers of normal seedlings were counted.

$$\text{Germination (\%)} = \frac{\text{Number of normal seedlings}}{\text{Total number of seeds}} \times 100$$

**Seedling parameters:** Root lengths and shoot lengths of ten seedlings were measured at 14 days after germination using the glass plate method in the laboratory with the help of a scale and graph paper and after that average was made out, expressed in centimetre (cm). A digital balance was used to measure the fresh weight of ten seedlings. After two hours of drying in a hot air oven at 80°C, the seedlings were weighed using a digital balance. The fresh weight and dry weight of the seedlings were both stated in grams (g).

**Vigour index:** Vigour index (VI) was computed by using the formula advised by Abdul-Baki and Anderson (1973):  $VI = G \times L$  Where, 'G' stands for germination percentage and 'L' denotes average seedling length (cm).

## 3. RESULTS AND DISCUSSION

### 3.1 Time of 50% Germination (Days)

The highest time to 50% germination over genotypes (6.529) was observed to produce by  $T_1$  on an average followed by  $T_2$ ,  $T_3$  and  $T_4$ ; while it was of shortest length for  $T_5$  preceded by  $T_6$  and  $T_7$ . Elkoca et al. (2007) observed that Osmo-priming by PEG solution improved time of 50% germination after seed treatment in pea. Highest time to 50% germination (5.725) was observed for  $G_1$  and lowest time to 50% germination was recognized for  $G_3$ , (4.020) over treatments (Table 1). When the interaction effect of genotypes and seed treatments were taken into consideration,  $G_1T_1$  showed highest value (7.753) for this parameter. Kundu and Bordolui (2023) found a similar result in carrots primed with Ag-nano particles.

### 3.2 Mean Germination Time (Days)

Treatments over genotypes, highest mean germination time was observed in  $T_1$  (7.922) followed by  $T_2$ ,  $T_3$  and  $T_4$ ; while it was minimum for  $T_5$  preceded by  $T_6$  and  $T_7$ . Hasan et al. (2016) found that Osmo-priming improved mean germination time after seed treatment in rice. Over treatments the highest mean germination

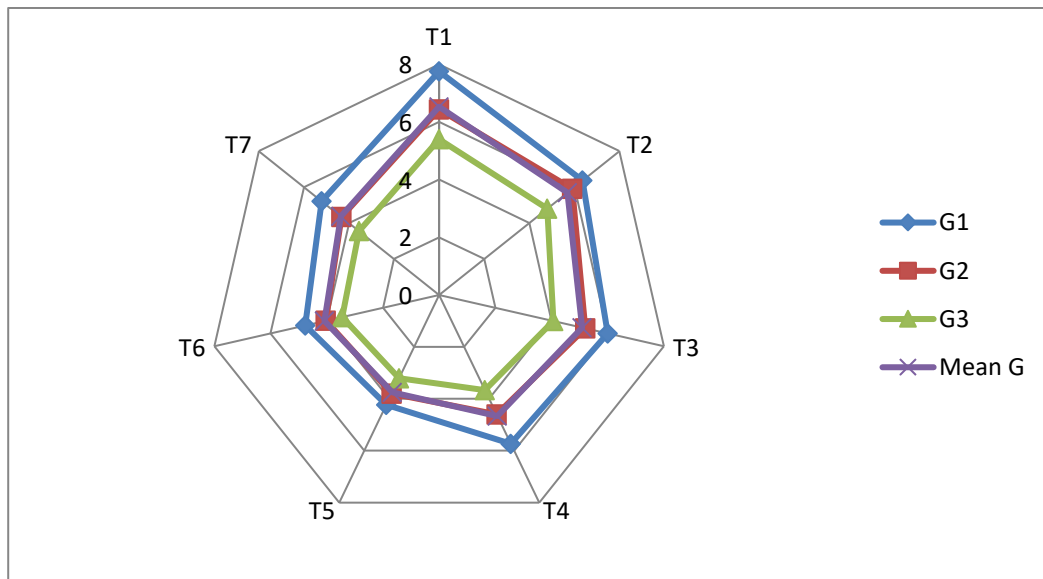
time was observed in G<sub>1</sub> (7.130) and lowest for G<sub>3</sub>, (5.407) (Table 2). G<sub>1</sub>T<sub>1</sub> showed highest value (9.147) for this parameter when interaction was made between genotypes and seed treatments.

But interaction value was non-significant. G<sub>2</sub>T<sub>1</sub>, G<sub>1</sub>T<sub>2</sub>; G<sub>3</sub>T<sub>2</sub>, G<sub>1</sub>T<sub>6</sub>; G<sub>3</sub>T<sub>3</sub>, G<sub>2</sub>T<sub>6</sub> were statistically at par. Ray and Bordolui found a similar result in tomatoes (2022a).

**Table 1. Effect of osmo-priming on Time of 50% Germination (days) of carrot genotypes**

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	7.753	6.367	6.000	5.733	4.233	4.767	5.220	5.725
G <sub>2</sub>	6.433	5.900	5.200	4.600	3.800	4.033	4.333	4.900
G <sub>3</sub>	5.400	4.797	4.067	3.667	3.207	3.450	3.550	4.020
Mean G	6.529	5.688	5.089	4.667	3.747	4.083	4.368	
	<b>G</b>		<b>T</b>		<b>GXT</b>			
SEm (±)	0.039		0.060		0.104			
LSD (0.05)	0.113		0.173		0.299			

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40MPa PEG-6000 for 48 hrs.



**Fig. 1. Graphical representation of Time of 50% Germination (days)**

**Table 2. Effect of osmo-priming on Mean Germination Time (days) of carrot genotypes**

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	9.147	7.793	7.393	7.147	5.660	6.160	6.613	7.130
G <sub>2</sub>	7.827	7.290	6.593	5.993	5.187	5.427	5.707	6.289
G <sub>3</sub>	6.793	6.140	5.460	5.060	4.623	4.860	4.910	5.407
Mean G	7.922	7.074	6.482	6.067	5.157	5.482	5.743	
	<b>G</b>		<b>T</b>		<b>GXT</b>			
SEm (±)	0.037		0.056		0.098			
LSD (0.05)	0.106		0.162		0.280			

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40MPa PEG-6000 for 48 hrs

### 3.3 Germination Index

Highest germination index over genotypes was observed in T<sub>5</sub> (5.503) followed by T<sub>6</sub>, T<sub>7</sub>, and T<sub>4</sub>, whereas T<sub>1</sub> (control) had the lowest germination index (2.089) preceded by T<sub>2</sub> and T<sub>3</sub>. Sadeghi et al. (2011) found that Osmo-priming by PEG solution improved time of germination index after seed treatment in soybean. Over the treatments, G<sub>3</sub> (4.922) had the highest germination index and G<sub>1</sub> (4.130) had the lowest (Table 3). When the interaction effect of genotypes and seed treatments were taken into consideration, G<sub>3</sub>T<sub>5</sub> (5.670) showed highest value for this parameter. But G<sub>1</sub>T<sub>1</sub> G<sub>2</sub>T<sub>1</sub>; G<sub>1</sub>T<sub>5</sub>, G<sub>2</sub>T<sub>7</sub> and G<sub>3</sub>T<sub>3</sub>; G<sub>3</sub>T<sub>4</sub>, G<sub>3</sub>T<sub>7</sub> were statistically at par.

### 3.4 Germination Energy (%)

The highest germination energy over genotypes was observed in T<sub>5</sub> (47.273) on an average followed by T<sub>6</sub>, T<sub>7</sub> and T<sub>4</sub>; while it was of lowest for T<sub>1</sub> (control) preceded by T<sub>2</sub> and T<sub>3</sub>. Sadeghi et al. (2011) found that Osmo-priming by PEG solution improved time of germination energy after seed treatment in soybean. Highest germination energy (42.450) was observed for G<sub>3</sub> and lowest germination energy was recognized for G<sub>1</sub> (40.256) over treatments (Table 4). When the interaction effect of genotypes and seed treatments were taken into consideration, G<sub>3</sub>T<sub>5</sub> showed highest value (47.687) for this parameter, though G<sub>1</sub>T<sub>3</sub> and G<sub>2</sub>T<sub>2</sub>; G<sub>3</sub>T<sub>7</sub> and G<sub>2</sub>T<sub>7</sub>; G<sub>2</sub>T<sub>6</sub>, G<sub>3</sub>T<sub>6</sub>; were statistically at par with each other.

### 3.5 Shoot Length (cm)

The longest shoot length over genotypes (3.759cm) was observed to produce by T<sub>5</sub>

followed by T<sub>6</sub>, T<sub>7</sub> and T<sub>4</sub>; while it was of shortest length for T<sub>1</sub> (control) preceded by T<sub>2</sub> and T<sub>3</sub>. Farooq et al. (2005) showed increased shoot length after seed treatment with Osmo-priming by PEG solution in rice. Highest shoot length (3.686 cm) was observed for G<sub>3</sub> and shortest shoot length (2.907 cm) was recognized for G<sub>1</sub>, over treatments (Table 5). Though G<sub>2</sub> and G<sub>3</sub> over treatment were non-significantly differ. When the interaction effect of genotypes and seed treatments were taken into consideration, G<sub>3</sub>T<sub>5</sub> showed highest value (4.392 cm) for this parameter, though G<sub>1</sub>T<sub>1</sub> and G<sub>2</sub>T<sub>1</sub>; G<sub>2</sub>T<sub>2</sub> and G<sub>3</sub>T<sub>2</sub>; G<sub>1</sub>T<sub>6</sub>, G<sub>1</sub>T<sub>7</sub> were statistically at par with each other. Similar outcomes were noted by Choudhury and Bordolui (2022a) in Bengal gram when they used sodium molybdate (Na<sub>2</sub>MoO<sub>4</sub>) nutri-priming to increase shoot length.

### 3.6 Root Length (cm)

T<sub>5</sub> (3.165 cm) produced highest root length (3.141 cm) over genotypes, followed by T<sub>4</sub>, T<sub>6</sub>, and T<sub>3</sub>, whereas T<sub>1</sub> (control) had the least root length, preceded by T<sub>2</sub> and T<sub>7</sub>. In pea, Yanglem et al. (2021) found that Osmo-priming by PEG solution improved root length after seed treatment. Over the treatments, G<sub>2</sub> had the highest root length (3.124 cm), and G<sub>1</sub> had the smallest root length (2.210 cm) (Table 6). Despite the fact that G<sub>1</sub> and G<sub>3</sub>, over treatments showed non-significant difference The interaction between genotypes and seed treatments G<sub>3</sub>T<sub>5</sub> showed highest value (3.165 cm) for this parameter, though G<sub>1</sub>T<sub>2</sub> and G<sub>2</sub>T<sub>2</sub>; G<sub>1</sub>T<sub>3</sub> and G<sub>1</sub>T<sub>4</sub> were statistically at par with each other. Choudhury and Bordolui (2022b) used potassium nitrate to observe a similar kind of result in Bengal gram.

**Table 3. Effect of osmo-priming on Germination index of carrot genotypes**

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	2.087	3.700	4.067	4.250	5.247	5.033	4.530	4.130
G <sub>2</sub>	2.057	4.633	5.153	5.213	5.593	5.377	5.250	4.754
G <sub>3</sub>	2.123	5.157	5.280	5.353	5.670	5.500	5.370	4.922
Mean G	2.089	4.497	4.833	4.939	5.503	5.303	5.050	
		<b>G</b>	<b>T</b>	<b>GXT</b>				
SEm (±)		0.016	0.025	0.044				
LSD (0.05)		0.047	0.072	0.125				

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40 MPa PEG-6000 for 48 hrs.

**Table 4. Effect of osmo-priming on germination energy (%) of carrot genotypes**

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	18.833 (25.707)	40.467 (39.488)	42.377 (40.599)	43.680 (41.353)	46.700 (43.091)	45.367 (42.324)	44.367 (41.749)	40.256 (39.187)
G <sub>2</sub>	21.667 (27.721)	42.167 (40.477)	42.967 (40.940)	45.167 (42.209)	47.433 (43.511)	46.333 (42.880)	45.333 (42.305)	41.581 (40.006)
G <sub>3</sub>	23.833 (29.208)	43.500 (41.248)	44.100 (41.595)	45.333 (42.305)	47.687 (43.656)	46.767 (43.139)	45.933 (42.650)	42.450 (40.542)
Mean G	21.444 (27.545)	42.044 (40.405)	43.148 (41.045)	44.727 (41.956)	47.273 (43.419)	46.156 (42.778)	45.211 (42.235)	
		<b>G</b>	<b>T</b>	<b>GXT</b>				
SEm (±)		0.087	0.133	0.230				
LSD (0.05)		0.249	0.380	0.659				

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40MPa PEG-6000 for 48 hrs.

**Table 5. Effect of osmo-priming on shoot length (cm) of carrot genotypes**

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	2.400	2.743	2.880	2.970	2.550	3.474	3.330	2.907
G <sub>2</sub>	2.503	2.925	3.090	3.585	4.335	3.726	3.774	3.420
G <sub>3</sub>	2.823	2.947	3.437	4.266	4.392	4.074	3.864	3.686
Mean G	2.576	2.872	3.136	3.607	3.759	3.758	3.656	
		<b>G</b>	<b>T</b>	<b>GXT</b>				
SEm (±)		0.037	0.057	0.099				
LSD (0.05)		0.107	0.164	0.284				

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40MPa PEG-6000 for 48 hrs.

**Table 6. Effect of osmo-priming on root length (cm) of carrot genotypes**

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	2.267	2.136	2.289	2.175	2.634	2.020	1.950	2.210
G <sub>2</sub>	2.523	3.193	3.247	3.381	3.623	3.057	2.844	3.124
G <sub>3</sub>	2.313	2.397	2.445	2.610	3.165	2.913	2.994	2.691
Mean G	2.368	2.575	2.660	2.722	3.141	2.663	2.596	
		<b>G</b>	<b>T</b>	<b>GXT</b>				
SEm (±)		0.041	0.062	0.108				
LSD (0.05)		0.117	0.178	0.309				

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40MPa PEG-6000 for 48 hrs.

### 3.7 Seedling Length (cm)

Among the treatments over genotypes, T<sub>1</sub> (control) observed shortest seedling lengths, which was preceded by T<sub>2</sub> and T<sub>3</sub>, while T<sub>5</sub> produced seedlings with highest length of 6.900 cm, followed by T<sub>6</sub>, T<sub>4</sub>, and T<sub>7</sub>. According to Singh et al. (2015), cowpea seeds treated with

Osmo-priming by PEG solution produced longer shoots. Here, G<sub>1</sub> and G<sub>2</sub> are non-significantly differ. According to Singh et al., cowpea seeds treated with Osmo-priming by PEG solution produced longer shoots (2014). G<sub>2</sub> had the longest shoots (6.544 cm) and G<sub>1</sub> had the shortest shoots (5.117 cm) over treatments (Table 7). When the interaction effect of

genotypes and seed treatments were taken into consideration, G<sub>3</sub>T<sub>5</sub> showed highest value (7.557 cm) for this parameter, though G<sub>3</sub>T<sub>1</sub> and G<sub>1</sub>T<sub>4</sub>; G<sub>1</sub>T<sub>3</sub> and G<sub>1</sub>T<sub>4</sub> were statistically at par with each other.

### 3.8 Germination Percentage

In case of treatments over genotypes significantly differ with each other. But T<sub>3</sub> recorded highest germination percentage (93.644), followed by T<sub>4</sub>, T<sub>2</sub>, and T<sub>6</sub> whereas T<sub>1</sub> (control) produced the lowest germination percentage preceded by T<sub>7</sub> and T<sub>5</sub>. T<sub>3</sub> and T<sub>4</sub> were non-significantly differing with each other. Lemmens et al. (2019) found that Osmo-priming by PEG solution improved germination percentage in wheat. Over the treatments, G<sub>1</sub> showed the lowest germination percentage (91.312) while G<sub>3</sub> had the highest germination percentage (92.126) (Table 8). Interaction between genotypes and seed treatments G<sub>3</sub>T<sub>3</sub> observed highest value (93.967). G<sub>1</sub>T<sub>1</sub>, G<sub>3</sub>T<sub>1</sub>;

G<sub>2</sub>T<sub>2</sub>, G<sub>3</sub>T<sub>2</sub>, G<sub>1</sub>T<sub>3</sub>, G<sub>3</sub>T<sub>2</sub>; G<sub>2</sub>T<sub>4</sub> and G<sub>2</sub>T<sub>5</sub> were statistically at par. Ray and Bordolui (2022b) discovered a similar kind of outcome in tomato.

### 3.9 Vigour Index

The highest vigour index over genotypes was observed in T<sub>5</sub> (639.032) followed by T<sub>6</sub>, T<sub>4</sub> and T<sub>7</sub>; while it was lowest for T<sub>1</sub> (control) preceded by T<sub>2</sub> and T<sub>3</sub>. Rouhi et al. (2010) found that Osmo-priming by PEG solution improved vigour index in clover. Highest vigour index (602.773) was observed for G<sub>2</sub> and lowest vigour index (467.527) was recognized for G<sub>1</sub>, over treatments (Table 9). Though G<sub>1</sub> and G<sub>3</sub> over treatment were non-significantly differ. When the interaction effect of genotypes and seed treatments were taken into consideration, G<sub>3</sub>T<sub>5</sub> showed highest value (693.447) for this parameter, though G<sub>2</sub>T<sub>1</sub> and G<sub>1</sub>T<sub>1</sub>; G<sub>1</sub>T<sub>2</sub>, G<sub>3</sub>T<sub>1</sub>; G<sub>1</sub>T<sub>4</sub> and G<sub>1</sub>T<sub>5</sub> were non-significant with each other.

**Table 7. Effect of osmo-priming on seedling length (cm) of carrot genotypes**

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	4.667	4.879	5.169	5.145	5.184	5.494	5.280	5.117
G <sub>2</sub>	5.027	6.118	6.337	6.966	7.958	6.783	6.618	6.544
G <sub>3</sub>	5.137	5.343	5.882	6.876	7.557	6.987	6.858	6.377
Mean G	4.943	5.447	5.796	6.329	6.900	6.421	6.252	
		<b>G</b>	<b>T</b>	<b>GXT</b>				
SEm (±)		0.060	0.091	0.158				
LSD (0.05)		0.171	0.261	0.453				

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40MPa PEG-6000 for 48 hrs.

**Table 8. Effect of osmo-priming on germination percentage of carrot genotypes**

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	85.583 (67.661)	91.300 (72.833)	93.367 (75.128)	93.400 (75.091)	93.533 (75.245)	91.867 (73.404)	90.133 (71.666)	91.312 (73.004)
G <sub>2</sub>	87.083 (68.910)	93.600 (75.323)	93.600 (75.380)	92.333 (73.904)	92.833 (74.444)	93.267 (74.933)	90.833 (72.356)	91.936 (73.607)
G <sub>3</sub>	85.917 (67.932)	93.933 (75.748)	93.967 (75.768)	93.867 (75.634)	91.767 (73.307)	93.267 (74.954)	92.167 (73.726)	92.126 (73.867)
Mean	86.194	92.944	93.644	93.200	92.711	92.800	91.044	
G	(68.168)	(74.635)	(75.425)	(74.876)	(74.332)	(74.430)	(72.583)	
		<b>G</b>	<b>T</b>	<b>GXT</b>				
SEm (±)		0.192	0.294	0.509				
LSD (0.05)		0.551	0.842	1.458				

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40MPa PEG-6000 for 48 hrs.

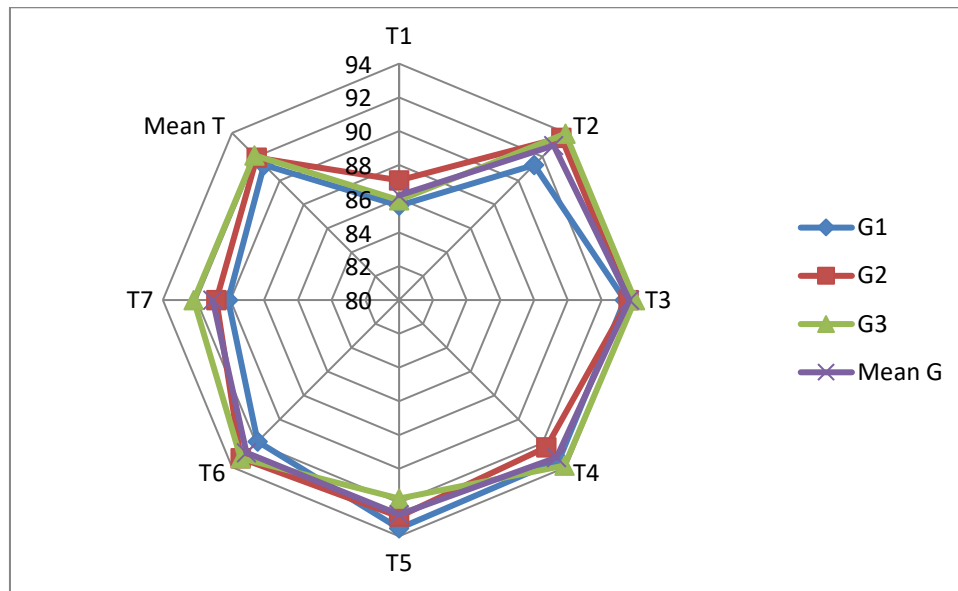


Fig. 2. Graphical representation of Germination (%)

Table 9. Effect of osmo-priming on vigour index of carrot genotypes

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	399.333	445.413	482.173	480.330	484.873	504.630	475.937	467.527
G <sub>2</sub>	437.747	572.743	593.107	643.340	738.777	632.587	601.113	602.773
G <sub>3</sub>	441.340	501.860	552.717	645.407	693.447	651.760	631.950	588.354
Mean G	426.140	506.672	542.666	589.692	639.032	596.326	569.667	
		G	T	GXT				
SEm (±)		5.390	8.234	14.261				
LSD (0.05)		15.438	23.582	40.845				

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40MPa PEG-6000 for 48 hrs

Table 10. Effect of osmo-priming on seedling fresh weight (mg) of carrot genotypes (10 seedlings)

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	62.333	49.000	76.000	87.333	102.000	95.000	86.000	79.667
G <sub>2</sub>	72.333	80.000	86.000	94.667	112.333	104.333	106.333	93.714
G <sub>3</sub>	74.667	79.000	82.667	92.000	108.333	103.000	103.333	91.857
Mean G	69.778	69.333	81.556	91.333	107.556	100.778	98.556	
		G	T	GXT				
SEm (±)		1.806	2.759	4.779				
LSD (0.05)		5.173	7.902	NS				

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40MPa PEG-6000 for 48 hrs.

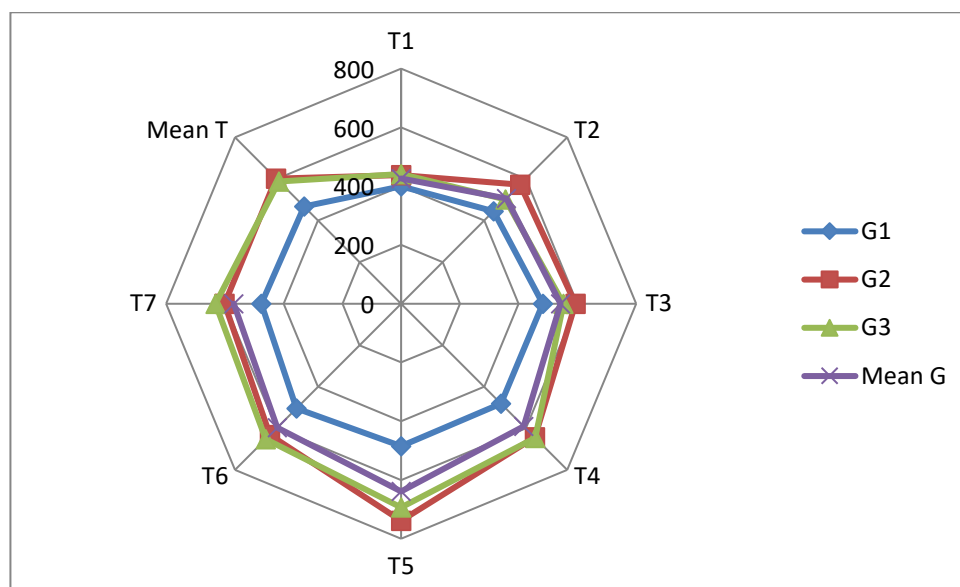


Fig. 3. Graphical representation of Vigour Index (%)

Table 11. Effect of osmo-priming on Seedling Dry Weight (mg) of carrot genotypes (10 seedlings)

	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	T <sub>4</sub>	T <sub>5</sub>	T <sub>6</sub>	T <sub>7</sub>	Mean T
G <sub>1</sub>	6.670	5.223	8.133	9.343	10.913	10.167	9.203	8.522
G <sub>2</sub>	7.740	8.560	9.203	10.130	12.020	11.163	11.380	10.028
G <sub>3</sub>	7.990	8.453	8.843	9.847	11.590	11.020	11.057	9.829
Mean G	7.467	7.412	8.727	9.773	11.508	10.783	10.547	
	<b>G</b>		<b>T</b>	<b>GXT</b>				
SEm (±)	0.193		0.295	0.511				
LSD (0.05)	0.553		0.845	NS				

Note: G = Genotypes, G<sub>1</sub> = Carrot Florence, G<sub>2</sub> = Deb Kuroda-1, G<sub>3</sub> = Deb Kuroda-3; T = Treatment, T<sub>1</sub> = Control, T<sub>2</sub> = 0.1 MPa PEG-6000 for 24 hrs, T<sub>3</sub> = 0.1 MPa PEG-6000 for 48 hrs, T<sub>4</sub> = 0.25 MPa PEG-6000 for 24 hrs, T<sub>5</sub> = 0.25 MPa PEG-6000 for 48 hrs, T<sub>6</sub> = 0.40 MPa PEG-6000 for 24 hrs, T<sub>7</sub> = 0.40 MPa PEG-6000 for 48 hrs.

### 3.10 Seedling Fresh Weight (mg) of 10 Seedlings

Over genotypes, T<sub>5</sub> produced the highest fresh weight (107.556 mg) followed by T<sub>6</sub>, T<sub>7</sub>, and T<sub>4</sub>; whereas T<sub>2</sub> showed the lowest fresh weight, preceded by T<sub>1</sub> and T<sub>3</sub>. Ghiyasi *et al.* (2008) found that Osmo-priming by PEG solution improved fresh weight after seed treatment in case of wheat. Similarly, Chakraborty and Bordolui (2021) discovered that Ag nano priming increased the fresh weight of green grams seedlings compared to other treatments. Genotypes over treatments, G<sub>3</sub> had the highest fresh weight (93.714 mg) and G<sub>1</sub> observed the lowest fresh weight (79.667 mg) (Table 10). When the interaction effect of genotypes and seed treatments were taken into consideration, G<sub>3</sub>T<sub>5</sub> showed highest value (108.333 mg) for this

parameter but they were non-significantly differ with each other.

### 3.11 Seedling Dry Weight (mg) of 10 Seedlings

The highest dry weight over genotypes was observed in T<sub>5</sub> (11.508) followed by T<sub>6</sub>, T<sub>7</sub>, and T<sub>4</sub>. But, T<sub>2</sub> showed the lowest dry weight, preceded by T<sub>1</sub> and T<sub>3</sub>. T<sub>1</sub> and T<sub>2</sub> were statistically at par. Ghiyasi *et al.* (2008) found that Osmo-priming by PEG solution improved dry weight after seed treatment in case of wheat. Over the treatments, G<sub>3</sub> had the highest dry weight (10.028), and G<sub>1</sub> had the lowest dry weight (8.522) (Table 11). G<sub>1</sub> and G<sub>2</sub> over treatments were non-significantly differing. The interaction effect of genotypes and seed treatments were non-significantly variation with

each other but G<sub>3</sub>T<sub>5</sub> showed highest dry weight (11.590).

#### 4. CONCLUSION

Carrot seeds treated with PEG-6000 had better seed quality than the control. In comparison to other treatments, PEG-6000 @ 0.25Mpa soaking for soaking duration 48 hours was the most effective treatment over genotypes. Significantly highest germination index, germination energy, germination percentage and lowest mean germination time were noted for Deb Kuroda-3 (G<sub>3</sub>) while highest seedling length, fresh weight, dry weight and vigour index were observed for Deb Kuroda-1(G<sub>2</sub>) although these genotypes were statistically at par. So, in germination point of view, Deb Kuroda-3 is best and in vigour point of view, Deb Kuroda-1 is best. For seed quality parameters such as germination energy (47.273), seedling vigour Index-I (639.032), and germination index (5.503), PEG-6000 @ 0.25Mpa soaking for 48 hours shown noticeably the best results. Consequently, PEG-6000 @ 0.25Mpa for soaking duration 48 hours is advised as a pre-sowing treatment for carrot seeds in order to improve seedling establishment.

#### DISCLAIMER (ARTIFICIAL INTELLIGENCE)

The authors of this manuscript hereby declare that no generative AI technologies, including text-to-image generators and Large Language Models (ChatGPT, COPILOT, etc.), were used in its writing or editing.

#### COMPETING INTERESTS

Authors have declared that no competing interests exist.

#### REFERENCES

- Abdul-Baki, A., & Anderson, J. D. (1973). Vigor determination in soybean seed by multiple criteria. *Crop Science*, 13(5), 630-633.
- Basu, R. N. (1976). Physico-chemical control of seed deterioration. *Seed Research*, 4(1), 15-23.
- Chakraborty, A., & Bordolui, S. K. (2021). Impact of seed priming with Ag-nanoparticle and GA3 on germination and vigour in green gram. *International Journal of Current Microbiology and Applied Sciences*, 10(3), 941-950.
- <https://doi.org/10.20546/ijcmas.2021.1003.119>
- Chakraborty, A., & Bordolui, S. K. (2021). Standardization of the appropriate doses of GA3 and Ag-nanoparticle in green gram for quality seed production. *International Journal of Environmental & Agriculture Research*, 7(4), 1-11.
- Choudhury, A., & Bordolui, S. K. (2022a). Seed invigoration treatment with sodium molybdate (Na<sub>2</sub>MoO<sub>4</sub>) nutri-priming for improvement of quality performance of Bengal gram (*Cicer arietinum* L.). *The Pharma Innovation Journal*, 11(12), 3381-3386.
- Choudhury, A., & Bordolui, S. K. (2022b). Inducement of seed priming with potassium nitrate on quality performance of chickpea (*Cicer arietinum* L.). *Biological Forum – An International Journal*, 14(4), 779-783.
- Coolbear, P., Francis, A., & Grierson, D. (1984). The effect of low temperature pre-sowing treatment on the germination performance and membrane integrity of artificially aged tomato seeds. *Journal of Experimental Botany*, 35(11), 1609-1617.
- Elkoca, E., Haliloglu, K., & Esitken, A. (2007). Hydro and osmopriming improve chickpea germination. *Acta Agriculturae Scandinavica Section B – Soil and Plant Science*, 57(3), 193-200.
- Ellis, R. H., & Roberts, E. H. (1981). The quantification of ageing and survival in orthodox seeds. *Seed Science and Technology*, 9, 373-409.
- Farooq, M., Basra, S. M. A., Ahmad, N., & Hafeez, K. (2005). Thermal hardening: A new seed vigor enhancement tool in rice. *Journal of Integrative Plant Biology*, 47(2), 187-193.
- Ghiyasi, M., Abbasi, S. E., Mehdi, T., Amirnia, R., & Hojat, S. (2008). Effect of osmopriming with polyethylene glycol (8000) on germination and seedling growth of wheat (*Triticum aestivum* L.) seeds under salt stress. *Research Journal of Biological Sciences*, 3, 91-94.
- Hasan, M. N., Salam, M. A., Chowdhury, M. M. I., Sultana, M., & Islam, N. (2016). Effect of osmopriming on germination of rice seed. *Bangladesh Journal of Agricultural Research*, 41(3), 451-460.

- Heydecker, W. (1973). Germination of an idea: The priming of seeds. *School of Agriculture Research, University of Nottingham*, 50-67.
- International Seed Testing Association (ISTA). (1996). *International rules for seed testing*. The International Seed Testing Association.
- Kundu, E., & Bordolui, S. K. (2023). Silver nanoparticles-mediated seed priming improves germination and physiological performance in carrot. *Biological Forum – An International Journal*, 15(10), 1079-1085.
- Lemmens, E., Deleu, L. J., De Brier, N., De Man, W. L., De Proft, M., Prinsen, E., & Delcour, J. A. (2019). The impact of hydro-priming and osmo-priming on seedling characteristics, plant hormone concentrations, activity of selected hydrolytic enzymes, and cell wall and phytate hydrolysis in sprouted wheat (*Triticum aestivum* L.). *ACS Omega*, 4(26), 22089–22100.
- Ray, J., & Bordolui, S. K. (2022a). Effect of seed priming as pre-treatment factors on germination and seedling vigour of tomato. *International Journal of Plant & Soil Science*, 34(20), 302-311.
- Ray, J., & Bordolui, S. K. (2022b). Seed quality deterioration of tomato during storage: Effect of storing containers and conditions. *Biological Forum – An International Journal*, 14(2), 137-142.
- Rouhi, H. R., Afshari, R. T., Moosavi, S., & Gharineh, M. H. (2010). Effects of osmo-priming on germination and vigour traits of bersim clover (*Trifolium alexandrinum* L.). *Notulae Scientia Biologicae*, 2(4), 59-63.
- Ruan, S., Xue, Q., & Tylkowska, K. (2002). The influence of priming on germination of rice (*Oryza sativa* L.) seeds and seedling emergence and performance in flooded soil. *Seed Science and Technology*, 30, 61-67.
- Sadeghi, H., Khazaei, F., Yari, L., & Sheidaei, S. (2011). Effect of seed osmo-priming on seed germination behavior and vigor of soybean (*Glycine max* L.). *ARPN Journal of Agricultural and Biological Science*, 6, 45-49.
- Singh, H., Jassal, R. K., Kang, J. S., Sandhu, S. S., Kang, H., & Grewal, K. (2015). Seed priming techniques in field crops – A review. *Agricultural Reviews*, 36(4), 251-264.
- Slama, I., Ghnaya, T., Hessini, K., Messedi, D., Savoure, A., & Abdelly, C. (2007). Comparative study of the effects of mannitol and PEG osmotic stress on growth and solute accumulation in *Sesuvium portulacastrum*. *Environmental and Experimental Botany*, 61(1), 10-17. <https://doi.org/10.1016/j.envexpbot.2007.02.004>
- Yanglem, S. D., & Ram, V. (2021). Effects of seed priming on root-shoot behavior and stress tolerance of pea (*Pisum sativum* L.). *Bangladesh Journal of Botany*, 50(2), 199-208.

**Disclaimer/Publisher's Note:** The statements, opinions and data contained in all publications are solely those of the individual author(s) and contributor(s) and not of the publisher and/or the editor(s). This publisher and/or the editor(s) disclaim responsibility for any injury to people or property resulting from any ideas, methods, instructions or products referred to in the content.

© Copyright (2025): Author(s). The licensee is the journal publisher. This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Peer-review history:

The peer review history for this paper can be accessed here:

<https://pr.sdiarticle5.com/review-history/130700>