



## Performance of *Cucurbita moschata* on Soil and Soilless Media

K. Okonwu<sup>1\*</sup>, M. I. Onyejanochie<sup>1</sup> and I. G. Ugiomoh<sup>1</sup>

<sup>1</sup>Department of Plant Science and Biotechnology, University of Port Harcourt, P.M.B. 5323, Port Harcourt, Nigeria.

### Authors' contributions

This work was carried out in collaboration between all authors. Authors KO and MIO designed the study and performed the statistical analysis. Authors KO and IGU wrote the protocol and wrote the first draft of the manuscript. Authors KO and IGU managed the analyses and literature searches of the study. All authors read and approved the final manuscript.

### Article Information

DOI: 10.9734/IJPSS/2018/v26i430050

#### Editor(s):

(1) Dr. Abhishek Naik, Technology Development Department - Vegetable Crops, United Phosphorus Limited -Advanta, Kolkata, India.

#### Reviewers:

(1) Paul Kweku Tandoh, Kwame Nkrumah University of Science and Technology, Ghana.

(2) Nusret Ozbay, Bingol University, Turkey.

Complete Peer review History: <http://www.sdiarticle3.com/review-history/46980>

Received 16 October 2018

Accepted 29 January 2019

Published 25 February 2019

Original Research Article

### ABSTRACT

*Cucurbita moschata* is widely grown in both tropical and temperate region due to its structural adaptability. The study was carried out to assess the performance of *C. moschata* on soil (humus) and soilless media (NPK 15:15:15 and NPK 20:10:15 growth media). These treatments are designated as T<sub>C</sub>, T<sub>A</sub> and T<sub>B</sub>, respectively. Standard procedures were followed in the assessment of mineral elements, nutritional composition, pigment compositions, and morphological characters (vein length, leaf area and number of leaves) of *C. moschata* in the three treatments. Among the treatments, *C. moschata* had the highest vein length, leaf area and number of leaves in T<sub>A</sub>, while T<sub>C</sub> recorded the least. Nutritional compositions of *C. moschata* were: moisture content (80.10%, 87.10% and 69.50%), carbohydrate (5.34%, 3.80% and 15.00%), ash content (3.61%, 1.20% and 4.10%), crude lipid (0.60%, 0.80% and 0.60%), crude protein (8.75%, 6.56% and 8.75%) and crude fiber (1.60%, 0.34% and 2.05%) for the treatments (T<sub>A</sub>, T<sub>B</sub> and T<sub>C</sub>). The mineral composition of *C. moschata* grown in T<sub>A</sub>, T<sub>B</sub> and T<sub>C</sub> growth media respectively were Mg (138.15 mg/kg, 43.90 mg/kg and 109.15 mg/kg), Mn (73.35 mg/kg, 0.25 mg/kg and 123.30 mg/kg), K (2,892.30 mg/kg, 3,338.80 mg/kg and 1,950.80 mg/kg), Zn (47.60 mg/kg, 10.55 mg/kg, 34.00 mg/kg), Ca (2,731.50 mg/kg,

\*Corresponding author: E-mail: [kalu.okonwu@uniport.edu.ng](mailto:kalu.okonwu@uniport.edu.ng);

337.95 mg/kg and 426.30 mg/kg), Na (89.65 mg/kg, 108.15 mg/kg and 66.60 mg/kg) and Fe (211.25 mg/kg, 0.00 mg/kg and 137.55 mg/kg) while copper was not detected. The pigment contents indicated the presence of chlorophyll *a* (0.32 mg/g, 0.39 mg/g and 0.24 mg/g), chlorophyll *b* (0.46 mg/g, 0.64 mg/g and 0.40 mg/g), carotenoid (0.33 mg/g, 0.42 mg/g and 0.30 mg/g), and xanthophyll (0.05 mg/g, 0.10 mg/g and 0.00 mg/g) for the three treatments, respectively. The study recommends the use of NPK 15:15:15 solution in a soilless condition and the inclusion of NPK 15:15:15 to the soil to improve the performance of *Cucurbita moschata*.

**Keywords:** Pumpkin; growth; development; minerals.

## 1. INTRODUCTION

The production of vegetables and food for human consumption by many subsistent farmers has largely depended on soil as the growth medium. This means of production accounts for majority of vegetables such as *Telfairia occidentalis*, *Cucurbita moschata*, and *Talinum triangulare* found in our local markets. The volumes of production of vegetables have declined in recent years in rural and urban areas due to anthropogenic activities and reduction in soil fertility. However, the advent of scientific research led to the cultivation of plants in a soilless medium like hydroponics. According to Kumar and Cho, hydroponic is a technology which aids plant growth in nutrient solution involving or excluding the application of external source for provision of mechanical support [1]. It was earlier reported by Jensen that production of food in soilless medium is on the increase all over the world [2]. In addition, hydroponically grown vegetables and fruits have been recorded in literature as possessing more nutritional and desirable values as compared to soil grown food produce [3,4,5,6]. The seedlings quality and vigour is dependent on the composition of media used [7,8,9,10]. Most research carried out on hydroponic has been geared towards leafy greens, peppers and tomato fruit [6,11,12], while research on hydroponically grown *C. moschata* has been scarce.

*Cucurbita moschata* (Duschene ex Lam.) Duschene ex Poir belongs to the family Cucurbitaceae. Cucurbitaceae ranks amongst the highest of plant families used as human food, cultivated in tropics and temperate regions [13]. *C. moschata* possesses nutritional and therapeutic qualities and has gained the attention of food scientists in recent time [14]. The seeds of *C. moschata* are rich in minerals [15], useful source of nutrients and oils [16] and thus could be used as valuable food supplement [15,17]. *C. moschata* as fruit vegetable is rich in carotenoids which have antioxidant activities and are easily

converted to retinol, the active form of vitamin A [18,19]. Beta carotene is the most predominant and active of the 5 or 6 provitamins present in commonly consumed foods [20]. It is locally consumed as freshly boiled and steamed or as processed food items in Thailand [21] and in cuisine or serve as desert in Malaysia [22]. There is also a wide variety carotenoid content of food from different races [23]. *C. moschata* is cultivated in Nigeria for both the fruits and leaves [24,25].

In line with the challenges of population dynamics round the globe and the reduction in arable land for the cultivation of plants, the study is aimed at evaluating the growth and development of *C. moschata* on both soil and soilless media and proffers information in order to enhance its production for both human consumption and profit making.

## 2. MATERIALS AND METHODS

The study was carried out within July – November 2017 and in a non-circulating hydroponics system [dimensions: 29 cm (width) x 41 cm (length) x 23 cm (depth)] in the screen house of Ecological Center, University of Port Harcourt.

### 2.1 Source of Materials Used

The seeds of *C. moschata* were collected from the Ecological Center of the University of Port Harcourt. The seeds were divided into two batches and planted in white-sand and humus soil, respectively. The medium of growth for the seedling were humus-soil ( $T_C$ ) and two NPK solution formulations (15:15:15 and 20:10:15) designated as  $T_A$  and  $T_B$ , respectively. The electrical conductivity, pH and total dissolved solutes of the solutions were 15.9  $\mu\text{s/cm}$ , 5.5, 10.18 ppm for NPK 15:15:15 and 14.8  $\mu\text{s/cm}$ , 5.3, 9.47 ppm for NPK 20:10:15, respectively. The two-week old seedlings raised with white-sand were transferred to hydroponic bowls

containing different solutions of NPK formulation ( $T_A$  and  $T_B$ ), which served as the soilless medium. The seedlings raised with humus soil served as the soil medium ( $T_C$ ). The plants were allowed to stand for 8 weeks after planting. The morphological characters of *C. Moschata* assessed were the vein length, number of leaves and leaf area. Minerals, pigment content and proximate composition of the leaves were determined following standard procedures.

## 2.2 Morphological Characters

Vein length of *C. moschata* was measured with meter rule calibrated in centimeters while the number of leaves was obtained by direct counting. The leaf area of *C. moschata* was determined using the method of Akoroda [26]. Estimated leaf area (LA) =  $0.9467 + 0.2475LW + 0.9724LWN$

Where,

- N = Number of leaflets in a leaf;
- L = Length of the central length;
- W = Maximum width of the central leaflet.

## 2.3 Proximate Composition

The proximate composition (crude protein, carbohydrate, crude fibre, crude lipid, ash and moisture contents) of *C. moschata* was determined using method of Association of Official Analytical Chemists [27].

## 2.4 Mineral Content

The mineral contents (Mg, Cu, Mn, K, Zn, Ca, Na and Fe) of *C. moschata* were determined using Atomic Absorption Spectrophotometer (AAS).

## 2.5 Pigment Content

Sample (0.1 g) *C. moschata* was transferred into a test tube and acetone was added to make it up to 10 ml. The test tube was then kept in the dark for 15 minutes with occasional shaking at room temperature. The chlorophyll, carotenoid and xanthophyll contents were analyzed spectrophotometrically by absorption measurement (A) at 350 nm to 700 nm with 1 nm interval and calculated according to the following equations:

$$\text{Chlorophyll } a \text{ (mg/g)} = \frac{13.7 \times A_{665} - 5.76 \times A_{649}}{\text{Mass} \times 200}$$

$$\text{Chlorophyll } b \text{ (mg/g)} = \frac{25.8 \times A_{649} - 7.6 \times A_{665}}{\text{Mass} \times 200}$$

$$\text{Carotenoid } \left(\frac{\text{mg}}{\text{g}}\right) = \frac{4.7 \times A_{440} - 0.263 \times \text{Chlorophyll (a + b)}}{\text{Mass} \times 200}$$

$$\text{Xanthophyll (mg/g)} = \frac{11.51 \times A_{480} - 20.61 \times A_{495}}{\text{Mass} \times 200}$$

The above pigments were extracted using acetone according to established methods [28,29,30].

## 2.6 Statistical Analysis

The data obtained for the morphological characters and pigment contents of *C. moschata* were subjected to statistical analysis using Microsoft Excel 2010 at 95% confidence level.

## 3. RESULTS AND DISCUSSION

### 3.1 Morphological Characters

#### 3.1.1 Vein length

The vein lengths of *C. moschata* grown in three different media are presented in Fig. 1. There was an increase in vein length from week 2 – 8 for the treatments. However,  $T_A$  medium gave the highest vein length (13.25 cm) compared to other treatments (11.15 cm and 9.10 cm) at 8<sup>th</sup> week. The least vein length (9.10 cm) was recorded in the soil treatment at 8<sup>th</sup> week.

#### 3.1.2 Leaf area

The leaf area of *C. moschata* grown in three different media is presented in Fig. 2. There was an increase from week 2 – 8 for the treatments,  $T_A$  treatment had the highest leaf area (74.35 cm<sup>2</sup>) compared to other treatments (67.05 cm<sup>2</sup> and 58.85 cm<sup>2</sup>) at 8<sup>th</sup> week. The least leaf area (58.85 cm<sup>2</sup>) was recorded in the  $T_C$  treatment at 8<sup>th</sup> week. This study has shown that the proportion of nitrogen, phosphorus and potassium available in the growth medium affects directly or indirectly the leaf area of plants.

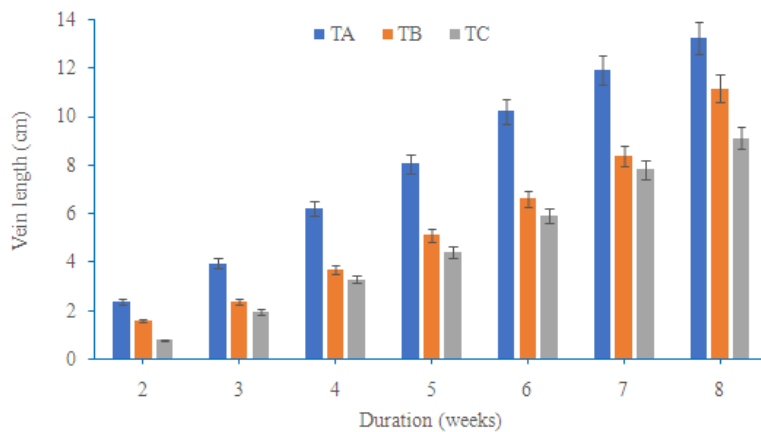


Fig. 1. Vein length (cm) of *C. moschata* in three different growth media

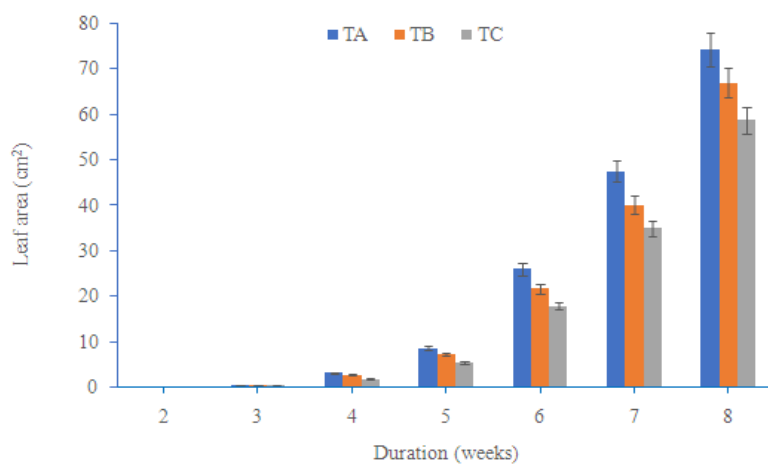


Fig. 2. Leaf area (cm<sup>2</sup>) of *C. moschata* in different growth media

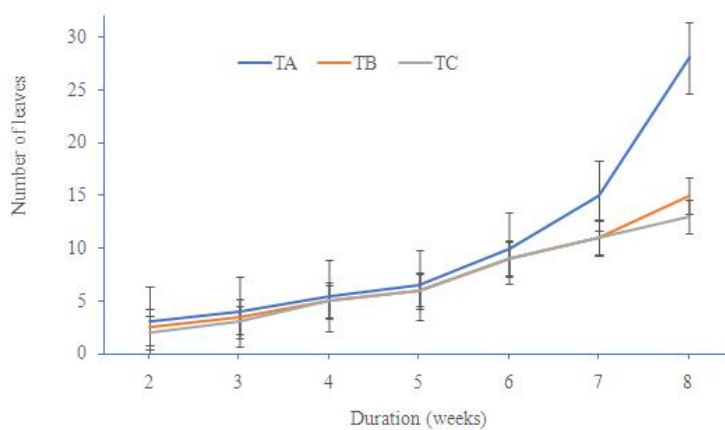
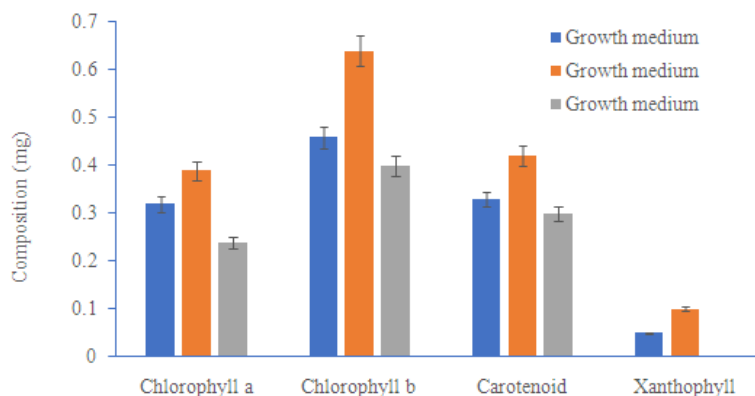


Fig. 3. Number of leaves of *C. moschata* in three different growth media



**Fig. 4. Pigment content (mg) of *C. moschata* in three different growth media**

### 3.1.3 Number of leaves

The number of leaves of *C. moschata* grown in three different media is presented in Fig. 3. From week 2 – 8, there was continuous increase in number of leaves among the three treatments ( $T_A$ ,  $T_B$  and  $T_C$ ). This observation is expected of growing plants. However, there was variation in the number of leaves of *C. moschata* in different treatments. At week 8, the highest increase in number of leaves was observed in  $T_A$  growth medium while the least recorded in  $T_C$  treatment. Apart from other factors that may interfere in plant growth, Nugawela et al. [31] reported a correlation between  $CO_2$  assimilation rate and planting conditions. Plants experiences reduced dry biomass and this affects vegetative growth due to the reduction in  $CO_2$  assimilation rate when planted under artificial shade such as green or shelter house. On the other hand, container and media interaction may affect fertility, pH, soluble salts, bulk density and root zone volume [32]. These may greatly influence plant growth on soilless substrate. However, the study has shown that the number of leaves of *C. moschata* was enhanced in soilless media containing varied proportion of NPK.

### 3.2 Proximate Compositions

The proximate composition of *C. moschata* leaves showed high amount of moisture content (80.10%, 87.10% and 69.50%) for  $T_A$ ,  $T_B$  and  $T_C$  treatments, respectively. The carbohydrate contents were 5.34%, 3.80% and 15.00% in that order. Others were: ash (3.61%, 1.20 % and 4.10%), protein (8.75%, 6.56% and 8.75%) and crude fibre (1.60%, 0.34% and 2.05%) were considerably low. The lipid contents (0.60%,

0.80% and 0.60%) were the lowest. The moisture and lipid contents were highest in *C. moschata* grown in  $T_B$  treatment. Ihenacho and Udebuani had earlier reported that high percentage moisture content provides for greater activity of water soluble enzymes and co-enzymes needed for metabolic activities [33]. Dietary fibre has some physiological effects in the gastro-intestinal tract such as: elimination of bile acids, fecal water [34]. It also serves as a source of human nutrition for diabetics in order to reduce glycaemic response to food and consequently the need for insulin [35]. Protein is an important part of catalytic activities, membrane build-up [36,37]. The nutrient composition of plant materials varies with season, environment, age and cultural practice [38].

### 3.3 Mineral Compositions

*Cucurbita moschata* leaves contain different minerals and their compositions ranging from lower concentrations of Zinc (Zn): 47.60 mg/kg, 10.55 mg/kg, and 34.00 mg/kg; Manganese (Mn): 73.35 mg/kg, 0.25 mg/kg, and 123.30 mg/kg; Sodium (Na): 89.65 mg/kg, 108.15 mg/kg, and 66.60 mg/kg; Magnesium (Mg): 138.15 mg/kg, 43.90 mg/kg and 109.15 mg/kg; Iron (Fe): 211.25 mg/kg, 0.00 mg/kg, and 137.55 mg/kg for  $T_A$ ,  $T_B$  and  $T_C$  treatments, respectively. Higher concentrations of mineral element were evident in Potassium (K): 2,892.30 mg/kg, 3,338.80 mg/kg, and 1,950.80 mg/kg respectively. In the three growth-media, Calcium (Ca) was highest in  $T_A$  medium (2,731.50 mg/kg). Copper content was not detected in the three growth-media. The role of these elements in the well-being of humans has been previously documented by previous workers [39]. Mineral element plays

diverse but essential role in plants, some of which include: catalytic, structural and electrochemical [37]. This implies that the consumption of *C. moschata* leaves will help to improve the nutritional status of human-beings.

### 3.4 Pigment Content

The leaves of *C. moschata* had the highest composition of chlorophyll *a* and chlorophyll *b* in T<sub>B</sub> medium, 0.39 mg and 0.64 mg respectively. The concentration of carotenoid and xanthophyll also had the highest concentration in T<sub>B</sub> medium, 0.42 mg and 0.10 mg respectively, as shown in Fig. 4. and higher than xanthophyll content in the growth media. Among the pigments, chlorophyll *b* content was in abundance than others in all the treatments. The carotenoids obtained in the leaves of *C. moschata* were in line with the work of Pritwani and Manthur, that reported the carotenoids value of 0.407 mg [40]. This trend could be associated with higher nitrogen content. Leaf growth, leaf area and photosynthetic rate may be influenced by the level of N in the soilless media. This ensures control of photosynthetic elements and production of carbohydrates. There may be probably a strong correlation and influence between chlorophyll and leaf area because the former indicates some level of N accumulation in leaves [41]. More so nitrogen use efficiency is said to be attributed to leaf area and other growth traits such as plant height [42]. Increase in N and P could increase leaf growth and chlorophyll content while its decrease may also be detrimental to crops [43]. Though, concentration of these elements sometimes may be advantageous or detrimental to the plants [44]. Other factors which may equally stimulate plant growth and development are better gaseous exchange; improved drainage and uniform extension of root systems sometimes are more advantageous than other growth factors [45].

### 4. CONCLUSION

*Cucurbita moschata* is rich with nutrient and mineral composition. The mineral composition of any growth medium determines the growth and development of *C. moschata*. The study has shown that the variation in the macro-nutrients affects the vigour of *C. moschata*. T<sub>A</sub> medium gave the highest vein length, leaf area, and number of leaves of *C. moschata* while the pigment compositions were slightly higher in T<sub>B</sub> medium compared to other treatments. The study therefore recommends that *C. moschata*

be grown in a moderate concentration of NPK solution with a view of tackling the problem of reduction in soil fertility and non-availability of arable land for the cultivation of *C. moschata*.

### COMPETING INTERESTS

Authors have declared that no competing interests exist.

### REFERENCES

1. Kumar RR, Cho JY. Reuse of hydroponic waste solution. Environmental Science and Pollution Research. 2014;1-9.
2. Jensen MH. Hydroponics worldwide. Acta Horticulturae. 1999;481:719-729.
3. Gichuhi PN, Mortley D, Bromfield E, Bovell AC. Nutritional, physical and sensory evaluation of hydroponic Carrots (*Daucus carota* L.) from different nutrient delivery systems. Journal of Food Science. 2009;74(9):403-412.
4. Sgherri C, Cecconami S, Pinzino C, Navari IZ, Izzo R. Levels of anti-oxidants and nutraceuticals in Basil grown in hydroponics and soil. Food Chemistry. 2010;123(2):416-422.
5. Selma MV, Luna MC, Martinez SA, Tudela JA, Beltran D, Baixauli C, Gil MI. Sensory quality of green and red fresh-cut lettuces (*Lactuca sativa* L.) are influenced by soil and soilless agricultural production systems. Post-harvest Biology and Technology. 2012;63(1):16-24.
6. Buchanan DN, Omaye ST. Comparative study of ascorbic acid and tocopherol concentrations in hydroponic and soil-grown lettuce. Food and Nutrition Sciences. 2013;4:1047-1053.
7. Corti C, Crippa L, Genevini PL, Centemero M. Compost use in plant nurseries, hydrological and physico-chemical characteristics. Compost Science Utilization. 1998;6:35-45.
8. Wilson SB, Stoffella PJ, Graetz DA. Use of compost as a media amendment for containerized production of two subtropical perennials. Journal of Environmental Horticulture. 2001;19(1):37-42.
9. Sahin U, Ors S, Ercisli S, Anapali O, Esitken A. Effect of pumice amendment on physical soil properties and strawberry plant growth. Journal of Central Europe Agriculture. 2005;6(3):361-366.
10. Baiyeri KP. Evaluation of nursery media for seedling emergence and early seedling

- growth of two tropical tree species. Journal of Agricultural Resources 2003;4(1):60-65.
11. Arias R, Lee TC, Specca D, Janes H. Quality comparison of hydroponic tomatoes (*Lycopersicon esculentum*) ripened on and off vine. Journal of Food Science. 2000;65(3):545-548.
  12. Koyama M, Nakamura C, Kozo N. Changes in phenols contents from Buckwheat sprout during growth stage. Journal of Food Science and Technology. 2013;50(1):86-91.
  13. Christenhusz MJ, Byng JW. The number of known plants species in the world and it's annual increase. Phytotaxa. 2010;261(3): 201-217.
  14. Fokou E, Achu M, Tchouanguep M. Preliminary nutritional evaluation of five species of Egusi seeds in Cameroon. African Journal of Food and Agriculture Nutrition Development. 2004;4:1-11.
  15. Nwofia GE, Nwogu N, Nwofia BK. Nutritional variation in fruits and seeds of pumpkins (*Cucurbita* spp) accessions from Nigeria. Pakistan Journal of Nutrition. 2012;11(10):848-858.
  16. Dhiman AK, Sharma KD, Attri S. Functional constituents and processing of pumpkin: A review. Journal of Food Science and Technology. 2009;46(5):411-417.
  17. Gorgonio CMS, Pumar M, Mothe CG. Macroscopic and physiochemical characterization of a sugarless and gluten-free cake enriched with fibres made from pumpkin seed (*Cucurbita maxima* L.) flour and corn starch. Gen. Tecnol. Aliment. Campinas. 2011;31(1):109-118.
  18. Dini I, Tenore GC, Dini A. Effects of industrial and domestic processing on antioxidant properties of pumpkin pulp. LWT- Food Science and Technology. 2003;53(1):382-385.
  19. Quiros ARB, Costa HS. Analysis of carotenoids in vegetable and plasma samples: A review. Journal of Food and Composition Analysis. 2006;19(2-3):97-111.
  20. McLaren DSM. Chapter 5 Vitamin A: The vitamins part 11 considering the individual vitamins. Elsevier. 2012;93-138.
  21. Pongjanta J, Naulbunrang A, Kawngdang S, Manon T, Thepjaikat T. Utilization of pumpkin powder in bakery products Songklanakarinn. Journal of Science and Technology. 2006;28(1):71-79.
  22. Norshazila S, Irwandi J, Othman R, Yumi HH. Carotenoid content in different locality of pumpkin (*Cucurbita moschata*) in Malaysia. International Journal of Pharmacy and Pharmaceutical Sciences. 2014;6(3):29-32.
  23. Bhaskarachary K, Rao DSS, Deosthale YG, Reddy V. Carotene content of some common and less familiar foods of plant origin. Food Chemistry. 1995;54:189-193.
  24. Okoli BE. Wild and cultivated cucurbits in Nigeria. Economic Botany. 1984;38:350-370.
  25. Ndukwu BC, Okoli BE. Studies on Nigeria *Cucurbita moschata*. Nigerian Journal of Botany. 1992;5:19-26.
  26. Akoroda MO. Non-destructive estimation of area and variation in shape of leaf lamina in the fluted pumpkin (*Telfairia occidentalis*). Scientia Horticulturae. 1993;53(3):261-267.
  27. AOAC. Official methods of analysis. Association of Official Analytical Chemicals, (15<sup>th</sup> Edition). Arlington, USA; 1990.
  28. Kukric ZZ, Topalic LN, Kukavica BM, Matos SB, Pavicic SS, Boroja MM, Savic AV. Characterization of anti-oxidant and microbial activities of nettle leaves (*Urtica dioica* L.). Acta Periodica Technologica. 2012;43:257-272.
  29. Chang SK, Nagendra PK, Amin I. Carotenoid retention in leaf vegetables based on cooking methods. International Food Research Journal. 2013;20(1):457-465.
  30. Duma M, Alsina I, Zeipiria S, Lepse L, Dubova L. Leaf vegetables as source of phytochemicals. Foodbaft. 2014;20:262-265.
  31. Nugawela A, Ariyawansa P, Samarasekara RK. Physiological yield determinants of sun and shade leaves of *Hevea brasiliensis*. Journal Rubber Research Institute Sri Lanka. 1995;76:1-10.
  32. Hockenberry MM, Cunliffe BA. Effects of media porosity and container size on overwintering and growth of ornamental grasses. HortScience. 2004;39(2):248-250.
  33. Ihenacho K, Udebuani AC. Nutritional composition of some leafy vegetable consumed in Imo State, Nigeria. Journal of Applied Science and Environmental Management. 2009;13(3):35-38.
  34. Akpabio UD, Akpan AE. Evaluation of nutritive and anti-nutritive composition of the seeds of *Mondora myristica* (African

- nutmeg). World Journal of Applied Science and Technology. 2012;4:49-55.
35. Onyije MO. Elemental and proximate contents of *Gnetum africanum* and *Telfairia occidentalis* (B.Sc. project). Department of Plant Science and Biotechnology, University of Port Harcourt. 2012;25-26.
36. Esenwo GJ. Developmental biology and plant physiology. Abeam Publishing Company, Nigeria. 2004;23-168.
37. Anoliefo CO. Introductory tropical plant biology. Uniben Press, Nigeria. 2006;257-362.
38. Apoxi SO, Long RJ, Castro FB, Orakor ER. Chemical composition and nutritive value of leaves and stems of tropical weeds. Grass and Forage Science. 2000;55(1):77-81.
39. Oluyemi EA, Akilua AA, Adenuya AA, Adebayo MB. Mineral contents of some commonly consumed Nigerian foods. Science Focus. 2006;11:153-157.
40. Pritwani R, Manthur P. Beta carotene content of some commonly consumed vegetables and fruits available in Delhi India. Journal of Nutrition and Food Science. 2017;7:625.
41. Nageswara RRC, Talwar HS, Wright GC. Rapid assessment of specific leaf area and leaf nitrogen in peanut (*Arachis hypogaea* L.) using chlorophyll meter. J. Agron. Crop Sci. 2001;189:175-182.
42. Hirel B, Le Gouis J, Ney B, Gallais A. The challenge of improving nitrogen use efficiency in crop plants: Towards a more central role for genetic variability and quantitative genetics within integrated approaches. Journal of Experimental Botany. 2007;58(9):2369–2387.
43. Sinclair TR, Vadez V. Physiological traits for crop yield improvement in low N and P environments. Plant and Soil. 2002;245:1-15.
44. Salisu M, Daud N, Ahmad I. Influence of fertilizer rates and soil series on growth performance of natural rubber (*Hevea brasiliensis*) latex timber clones. Australian Journal Crop Science. 2013;7:1998–2004.
45. Pinamonti F, Stringari G, Zorzi G. Use of compost in soilless cultivation. Compost Science & Utilization. 1997;5:38-46.

© 2018 Okonwu et al.; This is an Open Access article distributed under the terms of the Creative Commons Attribution License (<http://creativecommons.org/licenses/by/4.0>), which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

*Peer-review history:*

The peer review history for this paper can be accessed here:  
<http://www.sdiarticle3.com/review-history/46980>